idly and that adequate numbers of microorganisms are present in all samples to cause biodegradability. The initial inoculum is not critical as long as it is adequate to cause the lauryl sulfate to degrade in two days. Increasing the inoculum beyond this point does not seem to cause an appreciable increase in the rate of degradation and it does not cause a more complete degradation. Final levels of degradation are the same as in the standard river die-away test, with the accelerated test simply compressing the test time to less than one-half.

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[Received March 13, 1964—Accepted May 8, 1964]

Determination of the Glyceride Structure of Fats; Analysis of 14 Animal and Vegetable Fats¹

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Abstract

The glyceride compositions of seven animals and seven vegetable fats have been determined by GLC analysis of the oxidized esterified glycerides as described in an earlier paper in this series. The compositions determined are compared with those calculated from lipase hydrolysis data according to the method of VanderWal. Good agreement was found between the calculated and determined compositions for the majority of the 14 fats. The exceptions were human fat and the more saturated vegetable fats, palm oil and cocoa butter, where some discrepancies occurred.

Introduction

THE GLC ANALYSIS of oxidized esterified glycerides ↓ (1) provides a rapid and accurate means of determining the fatty acid distribution in natural fats. Initial investigations on a few fats showed good agreement between the determined glyceride composition and that calculated from lipase hydrolysis data according to the method of VanderWal (2). The present work extends this type of analysis to 14 animal and vegetable fats. The results obtained are compared with those calculated from lipase hydrolysis data.

Experimental

The vegetable oils studied were commercial, refined and bleached samples. The animal fats were coldextracted with chloroform from fresh adipose tissue. Twenty mg of each fat was oxidized with permanganate-periodate as described earlier (1). The resulting azelao-glycerides were methylated with diazomethane and the oxidized esterified glycerides analysed by GLC. Conditions for GLC analysis and methods of calculation were as previously described

The original fatty acid compositions of the fats were determined by GLC of their methyl esters on an o-phthalic-ethylene glycol column. The method of Youngs (3) was used for lipase hydrolysis in which the composition of the liberated fatty acids was determined rather than that of the unhydrolysed monoglycerides. All results are reported as mole percentages.

Results and Discussion

Tables I and II give the glyceride compositions as determined and as calculated from lipase hydrolysis data. Since the separation of the oxidized esterified

glycerides by GLC is dependent on their effective carbon number, those glycerides giving rise to the same carbon number emerge together and are determined as a group as indicated in the tables. The individual glycerides can be calculated from lipase hydrolysis data and this has been done. Comparisons are then made on the basis of the sum of the calculated glycerides in a group where two or more glycerides have the same carbon number. Since myristic acid is a minor component in the fats investigated, the calculated proportion of myristic-containing glycerides is small. Glycerides containing more than one myristic acid are less than 0.1%.

Tables III and IV give the original fatty acid compositions of the fats, as determined by GLC of their methyl esters, and those calculated from the glyceride compositions obtained. The agreement between these compositions for the individual fats serves as a check on the glyceride analysis. The fatty acid composition in the 1-3 positions is also given in the Tables. These latter figures were used in calculating the glycride compositions on the basis of VanderWal's theory.

Good agreement between the determined and calculated compositions was found for linseed, corn, olive, cottonseed and soybean oils. For the more saturated fats, cocoa butter and palm oil, the proportion of disaturated glycerides found was somewhat higher than that calculated, with the remaining glycerides being lower than calculated. In view of the good agreement obtained for the other vegetable fats, this suggests that the actual glyceride distribution for the more saturated vegetable fats may be slightly different than that predicted from lipase hydrolysis data. This however, requires further investigation.

Agreement between the calculated and determined glyceride compositions for the animal fats was generally good with the exception of human fat. In the latter case considerably more monosaturated glycerides were found than would be expected from lipase hydrolysis data, with a corresponding drop in the proportions of the fully unsaturated glycerides and more saturated glycerides. A similar pattern was found for two other samples of human fat. Since humans undoubtedly receive a much higher proportion of dietary fat than the other animals tested, this discrepancy may represent the effect of combined endogenous and exogenous fats.

In general it appears that glyceride composition calculated on the basis of lipase hydrolysis data provides a good estimate for the majority of natural

¹ Issued as NRC No. 8052. ² National Research Council of Canada Postdoctorate Fellow.

TABLE I

	G	lyceride Co	mposition	of Vegetab	le Fats					
Carbon No.	36	38	40	42	44	46	48	50	52	54
Possible glycerides	Uз	MU ₂	PU ₂ M ₂ U	$^{\rm SU_2}_{\rm MPU}$	P ₂ U MSU M ₂ P	PSU MP ₂ M ₂ S	${f SU_2} \\ {f P_3} \\ {f MPS}$	P ₂ S MS ₂	S ₂ P	Sa
Palm oil										
Found	5.9	0.9	30.5	7.6	35.6	8.8	9.1	1.3	0.3	
Calc.	9.4	1.1	29.9	6.1	30.1	9.1	9.8	3.8	0.5	
Linseed oil										
Found	73.0		17.3	8.0	1.0	0.6				
Calc	69.6		18,9	8.4	1.5	1.3	0.3			
Corn oil	00.0		10,0	0.2						
Found	60.5		30.0	4.3	4.0	1.1	0.2			
Calc.	60.2		31.1	3.8	4.4	0.3	0.3	0.1		
Cocoa butter	00.2		01.1	0.0	1.1	0.0	0.0	0.2		
Found	0.8		6.2	8.8	19.1	39.9	22.7	1.4	1.1	
Calc.	1.6	****	9.7	11.3	15.1	35.3	21.3	2.2	2.5	1.0
Olive oil	1.0	****	J. 1	11.0	10.1	00.0	-1.0			
	59.6		29.8	5.8	3.8	1.0			****	
Found	59.9	****	28.7	6.0	3.7	1.4	0.3		****	
Calc Cottonseed oil	35.5	****	20.1	0.0	0.1	***	0.0			
	27.1	2.0	42.4	6.5	18.0	3.0	1.0			
Found	26.6	1.5	43.6	5.8	18.8	3.0	0.7	****		
CalcSoybean oil	20.0	1.3	40.0	5.6	10.0	0.0	0.1		****	
	** *		000	0.0	4.8	2.3	0.2			
Found	55.5	****	28.0	9.2	3.3	$\frac{2.3}{2.4}$	0.6		• • • • •	
Calc.	56.5		27.2	10.0	3.3	4.4	0.0			

U-unsaturated acids; M-myristic acid; P-palmitic acid and S-stearic acid.

TABLE II

		Glyceride	Composition	on of Anima	l Fats					
Carbon No.	36	38	40	42	44	46	48	50	52	54
Possible glycerides	Uз	MU ₂	PU ₂ M ₂ U	$\begin{array}{c} SU_2 \\ MPU \\ M_3 \end{array}$	P ₂ U MSU M ₂ P	PSU MP ₂ M ₂ S	$\begin{array}{c} \mathrm{SU_2} \\ \mathrm{P_3} \\ \mathrm{MPS} \end{array}$	P ₂ S MS ₂	S ₂ P	Ss
Human										
Found	21.8	3.6	45.7	13.6	12.6	2.7				****
Calc	32.0	3.4	34.5	10.1	11.9	5.4	1.8	0.7	0.1	
Dog										
Found	20.3	4.2	24.8	20.7	14.1	10.2	4.4	0,9	0.4	
Calc.	22.8	4.4	25.8	17.1	11.7	10.8	4.2	2.0	0.8	****
Ground squirrel		•••	20.0							
Found	55.0	3.0	33,1	4.2	4.2	0.5				
Found Calc.	55.8	2.0	31.6	4.0	5.2	1.1	0.3			
Chicken	33.0	2.0	01.0	4.0	0.4		0,0		****	
	31.6	1.5	36.7	10,6	12.9	4.7	1.6	0.4		
Found			34.9	11.1	11.9	6.3	1.7	0.7	0.1	
Calc	31.7	1.6	34.9	11.1	11.5	0.0	1.1	0.1	0.1	
Pig			00.0	140	c 7	18.3	1.9	1.8	2.2	
Found	14.5	2.3	38.0	14.3	6.7				2.4	0.2
Caic	13.6	2.0	37.1	12.7	7.7	19.7	2.4	2.2	2.4	0.2
Rabbit										
Found	27.6	3.6	36.4	13.5	12.0	4.3	2.0	0.6		
Calc.	31.1	4.3	34.2	9.5	13.4	4.9	2.0	0.6		
Guinea pig										
Found	31.1		34.3	14.0	13.2	5.0	2.0	0.4		
Calc.	33.0		35.9	9.2	13.7	6.0	1.3	0.9		

U-unsaturated acids; M-myristic acid; P-palmitic acid and S-stearic acid.

TABLE III Fatty Acid Composition of Vegetable Fats

Fatty Acid Com	position	or vegetabl	e rats	
1	M	P	8	U
Palm oil				
Original	1.7	45.5	6.3	46.5
Calc	1.7	46.7	5.7	45.9
1-3 positions	2.2	51.9	6.5	39.4
Linseed oil	2.2	01.0	0.0	00.4
Original		7.8	3.4	88.8
Calc		6.8	2.9	90.3
1-3 positions	• • • • •		5.1	84.7
Corn oil	****	10.2	5.1	84.7
			• •	0.4 🖶
Original		13.5	1.8	84.7
Calc.	****	13.1	1.9	85.0
1-3 positions		17.7	2.5	79.8
Cocoa butter				
Original	****	28.1	32.9	38.3
Calc.		29.1	32.8	38.1
1-3 positions		40.1	46.8	13.1
Olive oil				
Original		12.6	2.6	84.8
Calc		12.7	2.3	85.0
1-3 positions		19.0	4.1	76.9
Cottonseed oil		13.0	4.1	10.5
Original	1.0	29.8	2.4	66.8
Calc.	1.1	28.5	2.8	
1-3 positions				67.6
	1.2	42.9	2.4	53,5
Soybean oil				
Original		12.2	4.4	83.4
Calc.		13.3	4.0	82.7
1-3 positions		18,1	6,6	74.7

U—unsaturated acids; M—myristic acid; P—palmitic acid and —stearic acid.

fats, particularly for those of commercial interest. Only the individual saturated acids have been considered here, the unsaturated acids being considered as a group. That the distribution of individual unsaturated acids may also be calculated in this way is indicated by a previous publication (4) where the distribution of each individual acid was determined for two fats.

TABLE IV
Fatty Acid Composition of Animal Fats

Fatty Acid Co	Composition of Animal Fats						
	М	P	S	U			
Human							
Original	2.4	23.4	5.2	69.0			
Calc	$^{2.0}$	25.3	4.6	68.1			
1-3 positions	2.4	28.5	6.8	62.3			
Dog							
Öriginal	4.2	22.8	11.7	61.0			
Calc.	3.8	24.0	11.3	60,9			
1-3 positions	2.7	22.9	16.5	58.2			
Fround squirrel							
Original	1.0	14.8	1.5	82.7			
Calc,	1.0	14.2	1.5	83.3			
1–3 positions	0.9	20.1	2.0	77.0			
Chicken							
Original	1.1	23.6	6.2	69.1			
Calc.	1.0	24.0	5.1	70.1			
1-3 positions	1.1	28.9	9.0	61.0			
Pig positions		20,0	0.0	00			
Original	1.5	26.0	12.5	60.0			
Calc.	1.0	25.8	13.8	59.4			
1-3 positions	0.7	8.2	18.9	72.2			
Rabbit	0.1	0,2	10.5	12.2			
Original	3.2	24.6	5.3	66.9			
Calc.	3.5	$\frac{24.0}{24.0}$	5.5	67.0			
1-3 positions	1.8	27.4	7.8	63.0			
	1,0	41.4	1.0	55.0			
Guinea pig		24.0	5.9	70.1			
Original	****	23.6	5.9 6.9	69.5			
Calc.				64.2			
1-3 positions		27.3	8.5	04.2			

U-unsaturated acids; M-myristic acid; P-palmitic acid and

ACKNOWLEDGMENTS

Technical assistance throughout the investigation by D. L. McPhee.

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[Received March 17, 1964—Accepted May 20, 1964]